# **Heart weight/tibia length ESLIM\_020\_001**

### **Purpose**

Relaxation, dissection, weighing and fixation of heart for histological analysis. Changes in heart weight and wall thickness are linked to cardiovascular phenotypes. This protocol describes the relaxation, dissection, weighing and fixation of the hearts to prepare the tissue for sectioning.



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#### 1.0 Purpose:

1.1 Relaxation, dissection, weighing and fixation of heart for histological analysis. Changes in heart weight and wall thickness are linked to cardiovascular phenotypes. This protocol describes the relaxation, dissection, weighing and fixation of the hearts to prepare the tissue for sectioning.

#### 2.0 Scope:

- 2.1 Individuals who have been trained, and are competent in performing the procedures described herein must follow this procedure.
- 2.2 Any queries, comments or suggestions, either relating to this SOP in general or to a specific problem encountered during a procedure, should be addressed to the leader of cardiovascular investigations.
- 2.3 Any deviances from this protocol must be reported to the leader of cardiovascular investigations.

#### 3.0 Safety Requirements:

3.1 General laboratory procedures should be followed, which include: no eating, no chewing gum, no drinking, and no applying of cosmetics in the work area. Laboratory coats and gloves must be worn at all times in the work area, unless the protocol specifically describes the appropriate attire for the procedure.

#### 4.0 Associated Documents:

- Tissue fixation with 4% buffered paraformaldehyde
- Trimming fixed tissues from necropsy
- Tissue processing and embedding in paraffin
- · Sectioning from paraffin embedded tissues
- Haematoxylin and eosin staining of histological sections



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- 5.1 Procedure is to be carried out on 3 males and 3 females at the end of pipeline 1.
- 5.2 The validity of results obtained from cardiovascular phenotyping is largely dependent on methods of animal husbandry. It is of vital importance that individuals following this procedure are experienced and aware of the animal's welfare, and is familiar with the animal being tested, in order to reduce the anxiety levels of the animal prior to testing.
- 5.3 The majority of mouse cardiovascular studies are age/sex/strain dependent. It is important to keep these parameters comparable throughout a single experiment.

#### 6.0 Quality Control:

6.1

#### 7.0 Equipment:

Fine forceps

Surgical scissors

Fine surgical scissors

"1 ml" syringes

"1.0 mm" needles

Centrifuge

 $\underline{\text{Tips}}$ 

Precision pipettes

"1.5 ml" tubes

Down draft table/fume hood certified for paraformade hyde use

Lab balance

Dissection tools

### 8.0 Supplies:

Ketamine (Imalgene®, Mérial)

Xylazine (Rompun® 2%, Bayer)

Sterile saline (0.9 % NaCl)



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8.1 250mM K.Cl

4% buffered paraformaldehyde

10ml syringe

26G butterfly needle connected to lucr valve (e.g. vacutainer blood collection kit)

7.5ml Bijou containers with lids

Labelling for bijous

C fold or equivalent paper towels

#### 9.0 Procedure:

Summary of protocol steps:

- Anaesthetize the mouse
- Intracardiac blood collection
- · Heart collection Collection of the heart and weight
- · Heart weight and storage for histology

#### 9.1 Anaesthe Anaesthetize the mousesia

- 9.1.1 Dilute ketamine and xylazine in sterile saline (NaCl 0.9%) in order to inject 150 mg/kg ketamine and 10 mg/kg xylazine to mice in 100 µL volume (for a 25 g-weight mouse).
- 9.1.2 Hold the mouse in your hand by the dorsal skin so that its head is up and its rear legs are down. Hold its tail with fingers.
- 9.1.3 Use "1mL" syringes and "0.5 mm" needles to inject anaesthesia solution and administrate 100 μL of solution per mouse (for a 25 g-weight mouse) by an intraperitoneal injection.
- 9.1.4 Check that mouse is deeply anaesthetized.

#### 9.2 Intracardiac blood collection

9.2.1 Lay the anesthetized mouse on its back.

9.2.2	Cut abdominal skin and wall to open peritoneal space.

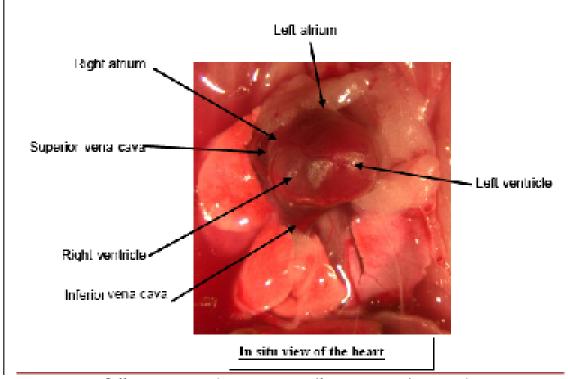
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- 9.2.3 Cut diaphra gm and ribs on both sides of the thoracic cage.
- 9.2.4 Pull up the thoracic cage.



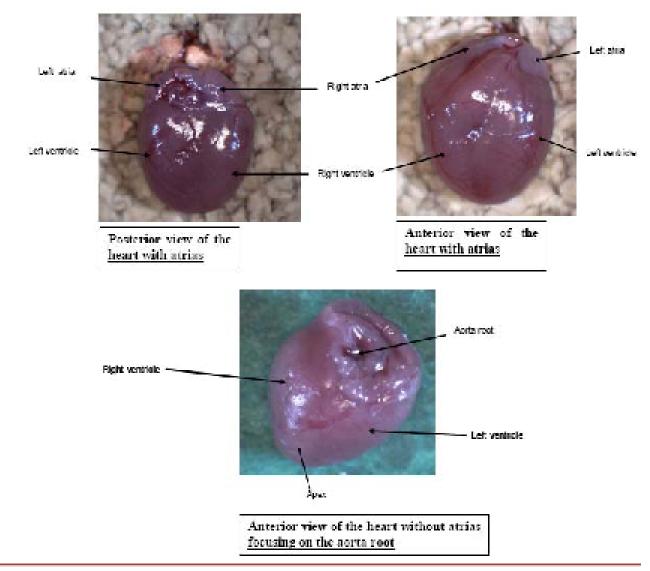
- 9.2.5 Carefully insert a "1.0 mm" needle connected to a "1 mL" syringe into the left ventricle and draw blood slowly in order that the wall does not collapse.
- 9.2.6 Transfer the blood into a "15.5 ml" tube coated with heparin.
- 9.2.7 Centrifuge blood (5000 rpm for 15 min at 4°C).
- 9.2.8 Collect the resulting supernatant (plasma) and store at –80°C until use.
- 9.3 Heart collection Collection of the heart and weight
- 9.3.1 Note if there is excessive fat surrounding the heart.
- 9.3.2 Remove the heart from the pericardiac membrane



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#### 9.3.4 Cut the major vessels at the point they enter/exit the atria and excise the heart.



9.3.5 9.3.5 ·

Label the bijous with the mouse ID

Load the K.Cl into the syringe, attach the butterfly needle and ensure all air bubbles are eleared.

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Record the anaesthesic type and dose

Once pedal reflex has ceased, but before the heart stops, open up the ventral abdominal cavity and expose the inferior vena cava below the liver and the ascending aorta above the heart.

Introduce K.Cl into the inferior vena cava. Once 0.5ml has been introduced, sever the earotid and perfuse another 5ml of K.Cl through the heart (or until the solution runs clear).

Note if there is excessive fat surrounding the heart.

Remove the heart from the pericardiac membrane

Note (text and photograph) if there are any abnormalities

Cut the major vessels at the point they enter/exit the atria.

Gently squeeze the heart empty and tap it onen the C-fold towels/surgical compress. D

and dab it dry to remove all remaining liquid in the heart.

- 9.3.6 Record the weight of the heart
- 9.4 Heart storage for histology (wax embedding)
- 9.4.1 Place the heart into 5ml of freshly made paraformaldehyde in a bijou and fix overnight at room temperature.
- 9.4.2 Embed the hearts in wax as per Tissue processing and embedding in paraffin
- 9.4.3 Trim the block from the atrial end until no more atria tissue is visible
- 9.4.4 Collect a single 5µm section and stain in H&E as described in Sectioning from paraffin embedded tissues and Haematoxylin and eosin staining of histological sections
- 9.4.5 Collect image of the whole section at 20x (minimum resolution of 0.5μm per pixel)
- 9.4.6 Note if there are abnormalities visible in the section
- 9.5 Alternative heart storage for histology (removal of atrias and cryopreservation and storage at -80°C)
- 9.5.1 Remove both atria. Then cut a slice of the ventricles. Fix tissues in 4% formalin



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9.5.2 Quickly freeze the apex in liquid nitrogen and store at -80°C for further investigation.

#### Parameters to recorded for each mouse

- Animal weight (g)
- Method of anaesthesia
- Anaesthetic dose
- Presence of excessive fat surrounding the heart (yes/no)
- Abnormal morphology of heart (yes/no) if yes, free text description
- Option of images taken to record abnormalities
- Heart weight (mg)
- Heart weight (mg)/tibia length (mm) from X-ray
- Other comments relating to the tissue collection
- Comments relating to the tissue and section processing
- Image of H&E stained section
- Comments on image

#### 10.0 Data Records and Reports:

10.1

#### 11.0 History Review:

11.1 Not applicable

#### 12.0 Emergency Procedures:

12.1 Ensure that appropriate steps are taken to eliminate the possibility of exposure to mouse allergens, needle sticks and paraformaldehyde poisoning.

#### **Parameters and Metadata**

### **Body weight** ESLIM\_020\_001\_001 | v1.0

simpleParameter

Req. Analysis: false Req. Upload: true Is Annotated: false

Unit Measured: g

**Description:** Body\_Weight

## **Heart weight** ESLIM\_020\_001\_002 | v1.0

simpleParameter

Req. Analysis: false Req. Upload: true Is Annotated: true

Unit Measured: mg

**Description:** Heart\_weight

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### **Tibia length** ESLIM\_020\_001\_003 | v1.0

simpleParameter

Req. Analysis: false Req. Upload: false Is Annotated: true

Unit Measured: mm		
Description: Tibia_length		
Visual abnormality simpleParameter	ESLIM_020_001_004   v1.0	
Req. Analysis: false	Req. Upload: false	Is Annotated: true
<b>Description:</b> Visual_abnorma	lity	
Options: yes, no,		
Description of visus simpleParameter	al abnormality ESLIN	/I_020_001_005   v1.0
Req. Analysis: false	Req. Upload: false	Is Annotated: false
<b>Description:</b> Description_of_\	visual_abnormality	

## **Image** ESLIM\_020\_001\_006 | v1.0

seriesMediaParameter

Req. Analysis: false Req. Upload: false Is Annotated: true

**Description:** Image Increments: Minimum 1 Histology abnormality ESLIM\_020\_001\_007 | v1.0 simpleParameter Req. Analysis: false Req. Upload: false Is Annotated: true **Description:** Histology\_abnormality Options: yes, no, Histology description ESLIM\_020\_001\_008 | v1.0 simpleParameter Req. Analysis: false Req. Upload: false Is Annotated: false **Description:** Histology\_description

## Histology image ESLIM\_020\_001\_009 | v1.0

seriesMediaParameter

Req. Analysis: false Req. Upload: false Is Annotated: true

**Description:** Histology\_image

### Pipeline number ESLIM\_020\_001\_801 | v1.0

procedureMetadata

**Increments:** Minimum 1

Req. Analysis: false Req. Upload: false Is Annotated: false

**Description:** Pipeline\_number

### Age of mice when culled ESLIM\_020\_001\_802 | v1.0

procedureMetadata

Req. Analysis: false Req. Upload: true Is Annotated: false

Description: Age\_of\_mice\_when\_culled

## Method of tibia measurement ESLIM\_020\_001\_803 | v1.0

procedureMetadata

Reg. Analysis: false Reg. Upload: false Is Annotated: false

**Description:** Method\_of\_tibia\_measurement

Options: xray, dissected tibia,

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### Available fixed ESLIM\_020\_001\_804 | v1.0

procedureMetadata

Req. Analysis: false Req. Upload: false Is Annotated: false

**Description:** Available\_fixed

Options: yes, no,

### Available embedded ESLIM\_020\_001\_805 | v1.0

procedureMetadata

Req. Analysis: false Req. Upload: false Is Annotated: false

**Description:** Available\_embedded

Options: yes, no,

### Histology performed ESLIM\_020\_001\_806 | v1.0

procedureMetadata

Req. Analysis: false Req. Upload: false Is Annotated: false

**Description:** Histology\_performed

Options: yes, no,

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